

Biological model for the study of the digoxin–quinidine drug interaction

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It is now well established that administration of the cardiac anti-arrhythmic drug, quinidine, can produce significantly elevated plasma levels of simultaneously administered digoxin in patients (Doering and König, 1978; Leahey et al., 1978, 1979). Although it has been estimated that this interaction may occur in up to 90% of patients receiving therapeutic doses of each drug (Bigger, 1979), the clinical implications of the elevated digoxin levels are unclear.

The chickens used in these experiments were mature, White Leghorn hens weighing 2–2.5 kg, and were housed in air-conditioned quarters with free access to food and water until the day before an experiment when the food was removed. The chicken was unanesthetized, but lightly restrained with a cloth harness that allowed it to remain comfortable in an upright position. A saphenous vein was cannulated using size no. 50 polyethylene tubing which was flushed continuously with 0.9% saline (0.05–0.1 ml/min). A three-way valve attached to the infusion line facilitated blood sampling and injections. Following cannulation the animal was again given free access to food and water. Room lights were adjusted to a 12-h cycle.

A single dose of about 350 ng [³H]digoxin was administered to 6 chickens by i.v. push over a one-minute period. This was followed by a flushing of the catheter with a small volume of saline after which the timed experiment was begun. In 9 experiments, quinidine sulfate was added to the constant saline infusion solution to result in an infusion rate of 180 µg/kg/h. This rate was calculated using human data for quinidine (therapeutic steady-state concentration, C_{av}^{ss} of 4 µg/ml; volume of distribution, V_{β} , of 0.5 ml · g⁻¹; terminal elimination rate constant β of 0.09 h⁻¹) (Bellet 1971; Guentert et al., 1979). The following two equations were used to calculate the loading dose, DL, and the infusion rate, R° :

$$R^{\circ} = C_{av}^{ss} \cdot V_{\beta} \cdot \beta \quad (1)$$

$$DL = C_{av}^{ss} \cdot V_{\beta} \quad (2)$$

In these experiments, the digoxin was injected i.v. one hour after starting the quinidine maintenance infusion. Venous blood samples were taken prior to digoxin dosing and at various times up to 72 h after dosing. A portion of each plasma sample was analyzed for radioactivity by standard techniques of liquid scintillation counting using a Beckman LS 7000 spectrometer and 3a70B cocktail (RPI, Mount Prospect, IL, U.S.A.). Results were expressed as ^3H cpm/ml plasma and converted to ng/ml.

To study the effect of quinidine on the renal digoxin clearance, serial urine samples (Sperber, 1948) were collected over a period of 180 min. Midway through each urine collection, a blood sample was drawn. A cross-over design was used by which digoxin clearance was first determined in the absence of quinidine and, 14 days later, during quinidine administration (2 mg/kg prime injection followed by 0.18 mg/kg/h maintenance infusion as described above). Clearance (ml/min) was calculated according to the following equation:

$$Cl_{\text{ren}} = \frac{[^3\text{H}]_{\text{urine}} \cdot \text{ml}_{\text{urine}}}{[^3\text{H}]_{\text{plasma}} \cdot \text{min}} \quad (3)$$

The digoxin drug concentration-time data were analyzed by a curve-fitting method (RESID program). All data sets could be fitted best to a biexponential model of the general equation:

$$C(t) = B \cdot e^{-\beta \cdot t} + A \cdot e^{-\alpha \cdot t} \quad (4)$$

The important pharmacokinetic parameters are listed in Table 1.

In addition a compartment model-independent analysis was performed based on the terminal disposition slope, the area under the curve by the trapezoidal rule and the dose administered. The volume of distribution by the area method, V_{area} , did not differ significantly from the V_{β} , and neither did the total clearance differ.

The mean digoxin blood concentration-time curves after i.v. push injection in the presence and absence of quinidine infusion are shown in Fig. 1. Significant differences between the two treatments exist at all time points between 1.5 and 30 h after i.v. digoxin injection.

Significant differences (see Table 1) were found for the volume of distribution at steady-state (V_{ss}), and the apparent volume of distribution, (V_{β} or V_{area}), the total clearance, (Cl_{tot}), and the area under the curve (AUC). The elimination half-life ($t_{1/2\beta}$) was practically identical in both groups. Since the clearance is the product of $V_{\beta} \times (0.693/t_{1/2\beta})$, the decrease in total clearance of digoxin in presence of quinidine seems to be caused predominantly or exclusively by the decrease in volume of distribution.

The renal clearance of digoxin decreased from 2.21 ml/min/kg in the absence of quinidine to 1.16 ml/min/kg, during quinidine infusion ($0.1 > P > 0.05$).

In spite of extensive research, the mechanism of the digoxin-quinidine drug interaction has not been fully elucidated. To be most meaningful, further studies should be carried out with pharmacokinetic digoxin disposition similar to that in

TABLE 1

PHARMACOKINETIC PARAMETERS OF DIGOXIN WITHOUT AND WITH COADMINISTRATION OF QUINIDINE

Parameter	Control (n = 6)			With quinidine (n = 9)			Unpaired <i>t</i> -test for S (<i>P</i> < 0.05)
	\bar{x}	\pm S.D.	\pm S.E.M.	\bar{x}	\pm S.D.	\pm S.E.M.	
Body weight BW (kg)	2.33	1.7	0.7	2.75	0.8	0.26	
Dose (ng)	368.7	50.9	20.8	344.5	46.0	15.4	
Distribution half-life							
$t_{1/2\alpha}$ (h)	0.34	0.25	0.10	0.55	0.85	0.28	N.S.
Elimination half-life							
$t_{1/2\beta}$ (h)	31.92	9.2	3.7	29.23	22.8	7.6	N.S.
Microconstants							
k_{12} (1/h)	2.16	1.10	0.45	2.06	1.49	0.50	N.S.
k_{21} (1/h)	0.22	0.05	0.02	0.21	0.12	0.04	N.S.
k_{13} (1/h)	0.27	0.13	0.05	0.93	1.5	0.49	N.S.
Volume of distribution							
V_c (l/kg)	0.91	0.71	0.29	0.41	0.33	0.11	N.S.
V_{ss} (l/kg)	7.79	2.90	1.20	2.82	1.91	0.64	S.
V_{β} (l/kg)	8.60	3.10	1.30	3.30	2.12	0.71	S.
Total clearance							
Cl_{tot} (ml/min/kg)	6.76	1.31	0.74	3.42	1.88	0.63	S.
Area under curve							
$AUC_{(0-\infty)}$ ((ng/ml)h)	0.944	0.36	0.15	1.89	0.88	0.29	S.

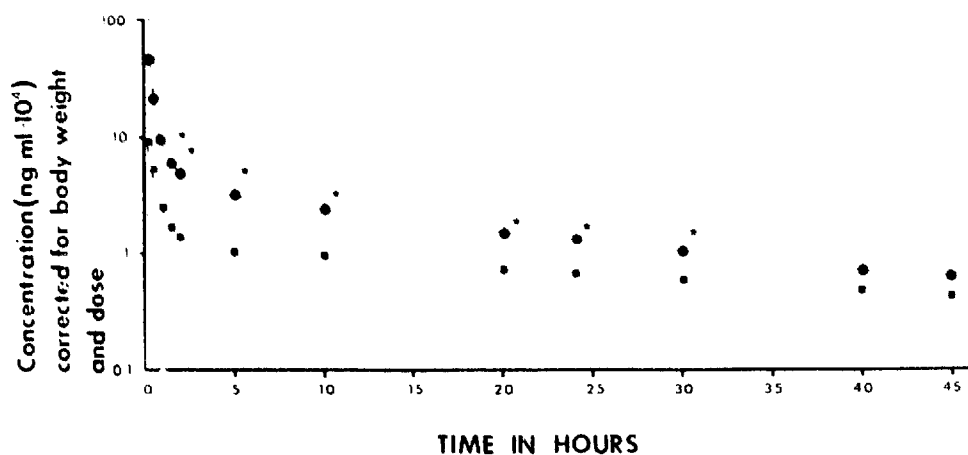


Fig. 1. Digoxin plasma concentration-time profile (means and S.D.) upon i.v. injection of digoxin alone (solid line) and in the presence of quinidine (dashed line). * indicates significant differences in concentration at the appropriate time (*P* < 0.05).

man. A comparison of the major pharmacokinetic parameters of digoxin in man with those obtained in various animal species is presented in Table 2. It is evident that the chicken may serve as a valid model for further studies. The i.v. injection of digoxin during i.v. infusion of quinidine was chosen in our studies because a change in digoxin absorption by quinidine has been ruled out as a significant contributor to the drug interaction (Hager et al., 1979; Hooymans and Merkus, 1978).

In most studies in man a reduction in the volume of distribution of digoxin has been observed (Schenk-Gustafsson et al., 1981; Leahey, 1980; Hager et al., 1979). Similar results have been obtained in the dog (Gibson et al., 1981). Our results are consistent with these findings. We found a reduction of V_B from 8.6 l kg^{-1} to 3.3 l kg^{-1} ($P < 0.005$). A decrease in the volume of distribution to 70% has been found by Hager et al. (1979), suggesting that quinidine may displace digoxin from tissue binding sites. In the present study using the chicken, we observed no significant difference in the digoxin volume of distribution of the central compartment in the presence or absence of quinidine, whereas V_{ss} and V_B were decreased significantly by quinidine. This indicates that the change in volume of distribution was associated with peripheral tissues. Where such a peripheral distribution volume change occurs needs further investigation. Results published to date have been contradictory. Whereas Straub et al. (1978) found that digoxin was displaced by quinidine from beef heart ATPase, no such displacement was observed by Doering (1979) in the

TABLE 2

PHARMACOKINETIC PARAMETERS OF DIGOXIN IN VARIOUS SPECIES \pm S.D. OR (RANGE)

Species	$t_{1/2\alpha}$ (h)	$t_{1/2\beta}$ (h)	V_c (l/kg)	V_{ss} (l/kg)
Man				
Adult	0.35 (0.27-0.50)	39.1 (26-60)	0.48 (0.24-0.78)	6.4 (4.8-8.1)
Newborn	0.59 (0.58-0.61)	49.3 (35-69)	1.1 ± 0.16	8.6 (7.5-9.9)
Infant	0.52 ± 0.1	29.8 ± 15.6	1.3 ± 0.16	9.96 ± 0.97
Elderly		69.6 ± 13.1		
Dog		25 (23-30)		9.11
Cat		57.8 ± 14.45		
Rat	0.25 (0.18-0.4)	2.5 ± 0.26	2.4 (1.9-2.9)	
Turkey	0.65 ± 0.63	11.03 ± 3.77	1.54 ± 0.54	4.56 ± 1.26
Chicken	0.34 ± 0.25	31.9 ± 9.2	0.91 ± 0.71	7.79 ± 2.9

sarcolemma fraction of lamb myocardium, or by Kim et al. (1981) in the Langendorff preparation of guinea pig heart. Also, in the presence of quinidine no change was observed in the amount of bound digoxin in kidney, cardiac muscle, liver, skeletal muscle and brain by the latter authors. Despite the lack of definitive data, the fact that digoxin does bind extensively to tissues, could mean that even minor decreases of binding in individual tissues could theoretically result in an overall effect of increased plasma concentrations.

Ochs and Greenblatt (1981) found a 64% decrease in Cl_{tot} of digoxin from a value of 6.06 ml/min/kg to 2.18 ml/min/kg in the presence of quinidine. In our chicken model, a similar decrease in Cl_{tot} was found from 6.76 ml/min/kg to 3.42 ml/min/kg (-51%).

The decrease in renal clearance of digoxin in presence of quinidine was first reported by Hooymans and Merkus (1978), who noted a decrease from 94 to 62 ml/min/1.73 m², or 34%. These results have been supported by studies in man (Doering, 1979; Hager et al., 1979; Leahey et al., 1980). Our studies in chickens demonstrated a decrease in digoxin renal clearance from 2.21 to 1.16 ml/min/kg, or 52%. The limited number of observations does not warrant a statistical evaluation; however, the results are qualitatively similar to observations in man.

Based on the comparison given above, we propose the chicken as a potential model for further digoxin-quinidine interaction studies.

V_{β} (l/kg)	Cl_{tot} (ml/min/kg) * (ml/min/1.73 m ²)	Cl_{ren} (ml/min/kg) * (ml/min/1.73 m ²)	References
6.8 (3-17)	5 (2.4-8) 188 ± 44 *	2.1 144 ± 4 ¹	Ochs et al., 1981; Koup et al., 1975; Turner et al., 1977; Lloyd et al., 1978; Reunig et al., 1973; Nyberg et al., 1974; Bonelli et al., 1978; Kramer et al., 1979; Rabkin et al., 1975; Gault et al., 1976; Ochs et al., 1979
11.1	3 45 (25-65) *		Wettrell, 1977; Morselli et al., 1975
12.1 ± 1.46	8.2 163 ± 98 *		Wettrell, 1977; Morselli et al., 1975; Dungan et al., 1972
4.1	0.8 ± 0.2		Cusack et al., 1979; Ewy et al., 1969
19 (17-21)	4.57		Doherty et al., 1966; Wilkerson et al., 1980
20.4 ± 5.34	4.05 ± 1.4		Weidler et al., 1978
3.5 (2.5-4.5)	5.77 ± 1.9	2.9	Harrison et al., 1976
5.55	5.81 ± 1.5		Park et al., 1981
8.6 ± 3.1	6.76 ± 1.81	2.21	present study

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